

23 the Tukey test at 5% of probability. Daily nutrient intake did not differ ($P>0.05$) between
24 the different treatments evaluated regardless of how it was expressed (kg/day or % body
25 weight [BW]), except for NDFap (% BW) that was higher ($P<0.05$) in the diet with
26 passion fruit residue than in diets with banana and mango residues. The digestibility of
27 ethereal extract was higher ($P<0.05$) for diets with sorghum silage, pineapple, and passion
28 fruit compared to the diet with banana residue. The digestibility of neutral detergent fiber
29 corrected for ashes and protein was higher ($P<0.05$) on animals receiving sorghum silage
30 than diets with mango and passion fruit residues. No difference ($P>0.05$) was observed
31 for purine derivative excretion, microbial efficiency, nitrogen intake, nitrogen loss (urine
32 and faeces), and nitrogen balance between diets. In conclusion, dehydrated fruit by-
33 products (pineapple, banana, mango, and passion fruit) are good options for partial
34 replacement of sorghum silage (75%) and potentially reduce feeding costs.

35 **Keywords:** Alternative feedstuff, Fruit residues, Lamb, Microbial nitrogen efficiency,
36 Nutritional value.

37 **1. Introduction**

38 Seasonal productivity of tropical forage is an important constraint to lamb production in
39 regions of tropical climate. Most production of forage occurs during the rainy season due
40 to greater availability of water and high temperatures, which in turn are essential for
41 tropical climate forages to express their potential growth (Silva et al. 2008). The opposite
42 is observed during the harmattern season, which is characterized by relatively low
43 temperatures and water precipitation, resulting in forage shortage. Thus, there is an
44 irregular production of forage throughout the years, and the use of by-products of fruit
45 processing as feed for sheep could be an alternative during the dry season.

46 Fruit production and processing has been growing, and so is the amount of by-products
47 produced. The use of agricultural technologies such as irrigation (Lousada Junior et al.
48 2005) has improved the production of fruits and promoted the expansion of fruit
49 processing agribusiness. A side consequence of this expansion has been the increasing
50 amount of residues produced over the years from different fruits (Matias et al. 2005),
51 since most part of the fruit is not used in industrial process. According to Waughon
52 (2006), about 77.5% of pineapple fruit production consists of bark, leaves, stems, crown,
53 and discarded fruits. About 40 to 60% of mango weight become residue after processed
54 for juice (Porrás 1989). Only 23.2% of passion fruit weight is used for juice production
55 (Ferrari et al. 2004).

56 Use of residues from fruit processing as sheep feed would reduce the environmental
57 impacts of fruit processing as well as reduce costs of animal feeding (Lousada Junior et
58 al. 2005). Ruminants have the unique ability to use fibrous materials as energy source due
59 to the physiological adaptation of their rumen. This means that the use of these feedstuffs

60 thereby could replace conventional feedstuff, reducing the competition between humans
61 and animals over these conventional feed ingredients.

62 Much of the residue produced is lost or misused due to the lack of knowledge about its
63 potential use. Precise estimates of dry matter (DM) intake is needed to promote efficient
64 nutrient intake in ruminants (NRC 2001). Physical-chemical characteristics of dietary
65 feedstuff and their interactions may have a great effect on dry matter intake (Allen 2000).
66 In addition, digestibility evaluation is essential to determine the nutritional value of
67 feedstuffs (Valadares Filho et al. 2000). According to Pereira (2003), knowledge about
68 nutrient digestibility of alternative feedings is essential to establish the energy they
69 supply.

70 Therefore, our study is aimed at evaluating intake, digestibility, microbial protein
71 production, and nitrogen balance of lambs fed with different dehydrated fruit residues
72 replacing sorghum silage.

73 **2. Materials and Methods**

74 All animal procedures were carried out in accordance with the guidelines described in the
75 Guide for the Care and Use of Agricultural Animals in Research and Teaching (FASS
76 2010). In addition, Ethics Committee on the Use of Animals from the Federal University
77 of the Jequitinhonha and Mucuri Valleys (UFVJM) approved the experimental
78 procedures used in this study (Protocol nº 018/2011 – UFVJM).

79 The experiment was conducted in the Ruminants Laboratory on the Experimental Farm
80 of Moura of the UFVJM, located in Curvelo – MG, Brazil at 18° 49' South latitude and
81 44° 23' West longitude. According to Köppen classification, the weather of the region is

82 Aw - Tropical Savanna with dry winter (Reboita et al. 2015). Bromatological analyses
83 were carried out in the Food Analysis Laboratory of the UFVJM – JK Campus,
84 Diamantina – MG, Brazil.

85 **2.1. Animals, diets, and handling**

86 Twenty-five non-castrated mixed breed Santa Inês and Texel male lambs with mean age
87 and body weights of 8 months and 20.64 ± 4.20 kg respectively were used for the study.
88 They were placed in individual metabolic crates of 1.5 x 1.0 m equipped with individual
89 feeding bunk and drinking fountain. The crates were cleaned on a daily basis. At the
90 beginning of the experiment, the animals were orally dewormed against endoparasites
91 using medicine with active ingredient of albendazole – 2 ml per 10 kg of live weight.

92 The experiment was carried out for 48 days between September and October of 2011,
93 consisting of 10-day acclimatization period, and 38 days for data collection. Lambs
94 received a total mixed ration formulated to have equal amount of protein and energy as
95 recommended by the NRC (2007) (Table 1).

96 We evaluated the inclusion of different fruit by-product as a substitute for sorghum silage
97 (dry matter basis). The animals were randomly assigned to one of the following five
98 treatments: (1) control, sorghum silage as forage source and no inclusion of fruit residue;
99 (2) pineapple, 75% replacement of sorghum silage for pineapple (*Ananas comosus* L.)
100 residue; (3) banana, 75% replacement of sorghum silage for banana (*Musa* spp.) residue;
101 (4) mango, 75% replacement of sorghum silage for mango (*Mangifera indica*) residue;
102 and (5) passion fruit, 75% replacement of sorghum silage for passion fruit
103 (*Passiflora* ssp.) residue. The forage to concentrate ratio of the diets were 40:60 (DM
104 basis) (Table 1) and were mixed together right before the animals were fed. The lambs

105 within each treatment were fed ad libitum diets twice a day (07:00 and 15:00). The
106 amount supplied to each animal was adjusted every day based on the leftovers of the
107 previous 24 hours allowing for approximately 10% of leftovers. Animals were provided
108 with water and mineral mixture ad libitum.

109 Throughout the study period, quantity of feed provided and quantity leftover as
110 unconsumed were daily recorded in order to measure individual consumption and to
111 adjust the amount supplied to ensure free-choice access. Diets and leftovers were daily
112 sampled as well, placed in plastic bags, and kept at - 10° C until analyses.

113 Different residues were composed of different parts of the fruit. Pineapple residue was
114 composed of the fruit skin and pressed pulp. Banana residue was composed of the fruit
115 peels. Mango residue was composed of the fruit peels, seeds, and pressed pulp. The
116 passion fruit residue was composed of the fruit peels and seeds. Upon arrival to the
117 experimental farm, residues were spread on top of a black plastic forming a 5 cm thick
118 layer. They were exposed to the sun for drying, being revolved three times a day, until
119 they show a dry and crumbly aspect. Then, they were ground on sieve No. 1 and used to
120 feed the animals. Nutrient composition of the residues and sorghum silage are shown in
121 Table 2.

122 Apparent digestibility of the nutrients was estimated on digestibility trials carried out
123 from days 30 to 34 of the experiment (five consecutive days). The total amount of feces
124 produced was collected using nappa leather (83% polyester and 17% cotton) bags adapted
125 into the animal's body. Before morning feeding, the feces produced by each animal were
126 weighed, sampled (about 10% of the total), and kept frozen at -20° C for later analyses.

127 Spot urine samples were used to quantify microbial protein production and nitrogen
128 balance. Urine samples were collected from day 35 to 37 at 08:00, 12:00, and 16:00h. We
129 attached a plastic bag to a galvanized wire loop and tied them to the back of the animals
130 using an elastic band. When spontaneous urination occurred, the bag was removed and
131 the urine sample was immediately processed. A sample of 10 ml of urine was measured
132 and diluted in 40 ml of 0.036 N sulfuric acid to prevent bacterial destruction of urinary
133 purine derivatives and precipitation of uric acid (Chen and Gomes 1992). Another 50 ml
134 sample was collected to quantify total nitrogen. Samples were labelled and stored at -15°
135 C for future analyses.

136 Blood was sampled in the jugular vein on the 38th day, approximately four hours after the
137 morning feeding using 8 ml tubes (Vacutainer Gel & Clot Blood Tube). Blood samples
138 were then centrifuged at 3,500 rpm for 15 minutes. Plasma was extracted, placed in plastic
139 tubes (Eppendorf® Safe-Lock Microcentrifuge Tube), and kept at -15° C for future
140 analyses.

141 **2.2. Chemical analyses and measurements**

142 Feedstuff (sorghum silage, fruit residues, and concentrate), leftovers, and faecal samples,
143 after drying, were grounded in a Wiley mill (Thomas Model 4 Wiley® Mill; Thomas
144 Scientific, Swedeboro, NJ, USA) to pass through a 1-mm stainless steel curved round-
145 hole sieve. Samples were analyzed for DM, organic matter (OM), crude protein (CP),
146 ethereal extract (EE), and ashes (AOAC 1997). Neutral detergent fiber (NDF) contents
147 were obtained using thermostable amylase (Termamy1120L,Novozymes) according
148 toMertens (2002). We used the Ankom® system for NDF evaluations, using bags (5.0 x
149 5.0 cm, porosity of 100 micrometers) which was made using non-woven fabric (TNT 100

150 g/m²). Acid detergent fiber (ADF), acid detergent insoluble nitrogen (ADIN), neutral
151 detergent insoluble nitrogen (NDIN), and lignin (LIG) (72% sulfuric acid) were obtained
152 by the sequential method of Robertson and Van Soest (1981) and presented as suggested
153 by Licitra et al. (1996).

154 Dry matter intake (DMI) was calculated based on the relationship between the DM
155 provided and the DM of the leftovers as follows: $DMI = (DM_{\text{diet}} \times \text{Quantity consumed}) -$
156 $(DM_{\text{leftover}} \times \text{Quantity of leftovers})$. Nutrient intake (NI) was calculated based on the
157 relationship between a nutrient and DM, and its content on diet and leftover as follows:
158 $NI = (\% \text{ Nutrient}_{\text{diet}} \times DM_{\text{ingested}}) - (\% \text{ Nutrient}_{\text{leftover}} \times DM_{\text{leftover}})$. Apparent digestibility
159 (AD) of the nutritional components from the diet was obtained using the equation
160 proposed by Silva and Leão (1979):

$$161 \quad AD = \frac{(DM_{\text{intake}} \times \% \text{nutrient}) - (DM_{\text{excreted}} \times \% \text{nutrient})}{DM_{\text{intake}} \times \% \text{nutrient}} \times 100$$

162 Non-fiber carbohydrates (NFC) were estimated according to Hall and Akinyode (2000):
163 $NFC = 100 - ([CP_{\text{total}} - CP_{\text{urea}} + \% \text{urea}] + NDF + EE + MM)$.

164 Total digestible nutrients (TDN) of the diet were calculated using the equation proposed
165 by the NRC (2001): $TDN = CPd + 2.25 \times EEd + NDFapD + NFC$; where TDN = total
166 digestible nutrients; CPd = digestible crude protein; EEd = digestible ethereal extract; and
167 NDFapD = neutral detergent fiber corrected for ashes and digestible protein; and NFC=
168 non-fibrous carbohydrates.

169 Creatinine and uric acid concentrations in urine as well as urea concentration in urine and
170 plasma were measured using a commercial kit (In Vitro®). Total volume of urine (TVU)
171 was estimated using the following equation (Chizzotti et al. 2006, Kozloski et al. 2005):

172
$$\text{TVU} = \frac{\text{BW} \times \text{Cre}_{\text{ref}}}{\text{Cre}},$$

173 where TVU is the total volume of urine produced (L); BW is the animal live body weight
174 (kg); Cre_{ref} is the referential daily excretion of creatinine (23 mg kg⁻¹ of BW); and Cre is
175 the concentration of creatinine on urine spot samples (mg l⁻¹). Urea nitrogen was obtained
176 by multiplying urea values by 0.4667. Purine derivatives (allantoin, xanthine, and
177 hypoxanthine) were determined in the diluted urine samples by the colorimetric method
178 proposed by Fujihara et al. (1987) and described by Chen and Gomes (1992). The total
179 excretion of purine derivatives (PD) was calculated by summing the results of allantoin,
180 uric acid, xanthine, and hypoxanthine excreted in the urine.

181 Absorbed microbial purines (Pabs) were calculated based on the excretion of purine
182 derivatives (EPD) in the urine using the equation proposed by Chen and Gomes (1992)
183 for sheep: $\text{EPD} = 0.84\text{Pabs} + (0.150 \text{BW}^{0.75} \exp^{-0.25\text{Pabs}})$, where $\text{BW}^{0.75}$ is the metabolic
184 BW of the animal. Pabs was estimated by solving the equation using the Newton-Raphson
185 iteration process (Chen and Gomes 1992).

186 The intestinal flow of microbial nitrogen (MN) was estimated based on Pabs using
187 equation proposed by Chen and Gomes (1992):

188
$$\text{MN} = \frac{\text{Pabs} \times 70}{0.116 \times 0.83 \times 1000}$$

189 Nitrogen balance (NB) was calculated as the difference between nitrogen intake and
190 nitrogen lost in urine and feces.

191 **2.3. Statistical analyses**

192 The experiment was conducted in a completely randomized design with five treatments
193 and five repetitions. Each animal was considered the experimental unit (n=5 per
194 treatment). The data were analysed using one-way analysis of variance and means were
195 compared using the Tukey test at 5% of probability. The analysis was done using SAS
196 version 9.0. The statistical model used was:

$$197 \quad Y_i = \mu + T_i + \varepsilon_i$$

198 where Y_i is the dependent variable (intake; feed digestibility; nitrogen balance etc.), μ is
199 the overall mean, T_i is the i th treatment (pineapple, banana, mango or passion fruit
200 residue; sorghum silage) effect, and ε_i is the residual error of the i th observation.

201 **3. Results**

202 The inclusion of dehydrated fruit by-products did not reduce the acceptability of the diets.
203 Treatments evaluated did not affect ($P>0.05$) the daily intake of DM, OM, CB, EE, NDF,
204 NDFap, NFC, NFCap, and TDN express in kg/day (Table 3). Treatments did not affect
205 ($P>0.05$) nutrients and TDN intake expressed as percentage of BW as well, except for
206 NDFap that was higher ($P<0.05$) among animals on passion fruit diet than animals on
207 banana and mango diets (Table 3).

208 The inclusion of pineapple, banana, mango, and passion fruit by-products as a partial
209 substitute to sorghum silage did not affect ($P>0.05$) the apparent digestibility coefficients
210 of DM, OM, CP, NFCap, and TDN (Table 4). The digestibility of EE was lower ($P<0.05$)
211 for banana diet compared to pineapple, passion fruit, and control diets. Animals on
212 passion fruit and mango diets showed lower ($P<0.05$) digestibility of NDFap compared
213 to the control diet.

214 The inclusion of fruit by-products as a partial substitute for sorghum silage did not
215 affected ($P<0.05$) urinary excretion of allantoin, xanthine-hypoxanthine, and uric acid
216 with average of 14.29, 2.34, and 1.02 mmol/day, respectively. The same was observed
217 for total PD and Pabs with an average of 17.62 and 20.318 mmol/day, respectively (Table
218 5). The proportion of allantoin, xanthine-hypoxanthine, and uric acid as a percentage of
219 the total was 80.65, 13.48, and 5.86%, respectively.

220 Sorghum silage replacement for dehydrated fruit residues did not affect ($P>0.05$)
221 microbial efficiency as well as urea nitrogen concentration in the urine ($P>0.05$).
222 However, animals on control diet had the highest ($P<0.05$) serum urea nitrogen, which in
223 turn was not different ($P>0.05$) from animals on banana residue diet (Table 5). Animals
224 on pineapple, mango, and passion fruit diets had about 33.09% lower serum nitrogen
225 concentration (average of 17.35 mg/dL) than animals on control diet (25.93 mg/dL).

226 The treatments evaluated did not affect ($P>0.05$) the intake and loss (feces and urine) of
227 nitrogen as well as nitrogen balance regardless of how it was expressed (g/day or g/kg
228 $BW^{0.75}$) (Table 6).

229 **4. Discussion**

230 We have carried out a study to evaluate the effect of partially replacing sorghum silage
231 with fruit by-products. The average dry matter intake DMI (1.13 kg/day; 3.46% BW) was
232 lower than expected, but was in agreement with the results found by other authors
233 (Lousada Junior et al. 2005, Vieira et al. 1999). NRC (2007) suggests that animals
234 weighing on average 20 kg would eat 1.2 kg of DM per day (6.0% BW). Lousada Junior
235 et al. (2005) evaluated the nutritive value of different fruit processing by-products for
236 sheep. They observed average DMI of 0.92 kg/animal and 1.20 kg/animal for diets with

237 pineapple and passion fruit by-products, respectively. The intake range in their study was
238 1.4 (Barbados cherry by-product) to 4.4% BW (guava by-product). These results are
239 similar to those found in our study.

240 We did not find differences in NDF intake in diets using dehydrated fruit by-products as
241 a substitute for sorghum silage. Sena et al. (2015), working with different levels of
242 substitution of Tifton 85 hay for passion fruit residue in diets for sheep, did not find
243 a significant difference for NDF intake between the treatments as well. They reported an
244 average NDF intake of 0.61 kg/day, which was higher than what we found (0.27 kg/day).
245 Diets with high NDF content are not desirable, because high NDF may result in low DMI
246 due to the physical filling effect of the rumen (Mertens 1994).

247 A possible explanation for the highest consumption (% BW) of neutral detergent fiber
248 corrected for ashes and protein (NDFap) on control, passion fruit, and pineapple diets
249 would be the concentration of NDFap on these feedstuffs as well as the characteristic of
250 their fiber. In general, fruit residues show high variation on their NDFap content;
251 Therefore, using only their NDF content to explain animals' feed intake as proposed by
252 NRC (2007) would not be appropriate for animals fed fruit by-products. The variation of
253 NDFap composition is the result of differences in cell wall composition. The proportion
254 of each cell wall component influences the intake of NDFap, mainly because they
255 influence its digestibility that in turn affect nutrient intake.

256 The lower digestibility of NDFap on passion fruit diet compared to the control diet could
257 be explained by the high lignin content of this by-product. The presence of lignin tends
258 to increase the indigestible fraction of the fiber, reducing the potentially digestible
259 fraction (Wilson 1994).

260 The average TDN digestibility (67.01%) from our study was higher than reported by
261 Lousada Junior et al. (2005) for diets with pineapple (45.6%) and passion fruit (52.9%)
262 residues. We believe it was due to differences in the by-product composition, especially
263 the pineapple residue because the crown was not included as part of the by-product in our
264 study. In addition, the intake of TDN (0.85 kg/ day; 2.62 % BW) was not affected by the
265 inclusion of any fruit by-products evaluated in our study. Altogether, these results
266 indicated that the use of fruit residues as a partial replacement for sorghum silage would
267 not impair the intake and digestibility of TDN.

268 The results of PD found in our study were similar to other studies. Allantoin was the most
269 abundant PD similar to Ma et al. (2013). They reported urinary allantoin values of 14.36,
270 10.24, and 6.33 mmol/day for *ad libitum*, 70%, 50% feed intake treatments, respectively.
271 They also reported values of xanthine-hypoxanthine ranging from 1.54 to 0.98 mmol/day
272 and uric acid ranging from 2.52 to 1.73 mmol/day, respectively, for *ad libitum*, 70%, and
273 50% feed intake treatments. In our study, we found an average of 14.29, 2.34, and 0.98
274 mmol/day, respectively, for allantoin, xanthine-hypoxanthine, and uric acid.

275 The flow of microbial nitrogen (average of 12.50 g/day) found in our study was lower
276 than reported by Fonseca et al. (2006), who observed values ranging from 15.7 to
277 20.6 g/day for dairy goats, but it was within the range of 14.2 to 8.0 g/day as reported by
278 Santos et al. (2016) for lambs.

279 Microbial efficiency observed in our study was lower than established recommendations.
280 Valadares Filho et al. (2006) proposed the value of 120 g microbial N/kg TDN for cattle
281 raised under tropical weather condition while the NRC (2001) establishes the value of
282 130 g microbial N/kg TDN. In our study, however, we found an average of

283 93.92 g microbial N/kg TDN which was lower than recommended as well as lower than
284 the average of 115.82 g microbial N/kg TDN reported by Silva et al. (2016).

285 Serum urea nitrogen of animals receiving control diet was higher than animals receiving
286 experimental diets with pineapple, mango, and passion fruit residues. Ruminal
287 microorganisms degrade about 50 to 70% of the CP that reaches the rumen and release
288 ammonia as the result. When ruminal ammonia concentration exceeds the
289 microorganisms' capability of metabolization, it is absorbed and reaches the liver where
290 is transformed in urea. Hence, plasma urea concentration it is the result of urea
291 synthesized in to the liver as well as the urea from amino acid metabolism. Part of this
292 urea is recycled and returned to the rumen through the saliva or absorbed through the
293 rumen wall, while the remaining is excreted in the urine (Kozloski 2002). Our results
294 suggest that animals fed with residues made better use of the nitrogen from the diet as
295 opposed by those that received sorghum silage.

296 Nitrogen balance was not different between experimental diets evaluated, which indicates
297 the animals retained protein from the diets, achieving the main objective of the nutritional
298 planning. When protein degradation rate exceeds carbohydrate fermentation, large
299 amount of nitrogen compounds can be eliminated through the urine and low performance
300 is observed due to unbalanced energy and protein intake as well as hepatic production of
301 urea (Van Soest 1994). Therefore, our results indicated that the experimental diets
302 provided a balanced supply of protein and energy, which in turn may have improved the
303 use of dietary protein.

304 In a conclusion, dehydrated fruit processing by-product (pineapple, banana, mango, and
305 passion fruit) could be used as a partial replacement of sorghum silage (75% DM basis)

306 to feed mixed-breed lambs since it did not affect nutrient intake (kg/day), microbial
307 protein synthesis, purine derivative excretion, and nitrogen balance. In addition, they are
308 a good alternative during forage shortage and their use could potentially reduce feeding
309 costs.

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316 **Statement of Animal Rights**

317 All procedures performed in studies involving animals were in accordance with the
318 ethical standards of the institution or practice at which the studies were conducted.

319 **Conflict of interest Statement**

320 The authors have no conflict of interest to declare.

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Table 1. Ingredients and chemical composition of the experimental diets¹.

Item ²	Treatment				
	Pineapple	Banana	Mango	Passion fruit	Sorghum silage
	30	30	30	30	40
<i>Ingredients (g kg⁻¹)</i>					
Sorghum silage	10	10	10	10	-
Ground corn	30	40	25	37	43
Soybean meal	15	18.5	18.5	13	15
Wheat bran	13.5	-	15	8.5	-
Urea/Ammonium sulfate (9:1)	-	-	-	-	0.5
Mineral premix ³	1.5	1.5	1.5	1.5	1.5
<i>Composition</i>					
DM (g kg ⁻¹)	86.4	86.4	86.9	87.1	71.2
<i>g kg⁻¹ (DM basis)</i>					
OM	92.5	90.7	92.9	88.1	91.5
CP	16.8	17.4	18.1	16.9	17.3
NDIN	12.6	28.8	13.4	14.9	16.7
ADIN	6.8	14.6	6.4	10.2	9.8
EE	2.6	4.2	3.7	3.5	2.8
Ashes	7.4	9.3	7.1	11.9	8.4
TC	73.0	69.7	71.4	67.9	71.4
NDF	27.7	27.3	26.0	30.8	30.2
NDFap	21.9	19.1	17.9	26.0	24.3
NFC	45.0	41.9	45.0	35.8	41.2
NFCap	50.9	50.0	53.2	41.6	47.1
ADF	13.1	11.6	11.8	16.8	13.4
HEM	14.5	15.6	14.2	14.0	16.8
CEL	10.9	7.9	9.1	11.3	10.9
LIG	2.0	3.5	2.5	4.9	2.4

¹Experimental diets consisted of sorghum silage (control diet) and 75% replacement of sorghum silage for dehydrated fruit processing by-products.

²DM = Dry matter; OM = organic matter; CP = crude protein; NDIN = neutral detergent insoluble nitrogen ($g\ kg^{-1}$ of total nitrogen); ADIN = acid detergent insoluble nitrogen ($g\ kg^{-1}$ of total nitrogen); EE = ethereal extract; TC = total carbohydrates; NDF = neutral detergent fiber; NDFap = neutral detergent fiber corrected for ashes and protein; NFC = non-fibrous carbohydrate; NFCap = non-fibrous carbohydrate corrected for ashes and protein; ADF = acid detergent fiber; HEM = hemicellulose; CEL = cellulose; LIG = lignin.

³The premix contained (per kg): 147 g Na, 120 g Ca, 87 g P, 18 g S, 3800 mg Zn, 1800 mg Fe, 1300 g Mn, 870 mg F, 590 mg Cu, 300 mg Mo, 80 mg I, 40 mg Co, 20 mg Cr, and 15 mg Se.

Table 2. Nutrient composition of sorghum silage and dehydrated fruit processing by-products.

Item ¹	Pineapple	Banana	Mango	Passion fruit	Sorghum silage
DM	91.8	90.7	92.2	93.6	41.1
<i>g kg⁻¹ (DM basis)</i>					
CP	11.0	12.0	9.5	12.0	6.2
NDIN	25.7	81.6	26.9	28.6	36.7
ADIN	6.8	14.1	6.4	10.2	9.8
EE	2.2	7.5	4.7	3.8	2.6
Ashes	6.5	16.4	3.1	13.2	5.3
TC	80.5	66.3	83.7	72.0	86.0
NDF	50.6	44.3	33.7	53.4	57.5
NDFap	46.1	35.1	25.5	52.0	52.8
NFC	28.7	20.2	49.0	13.8	28.5
NFCap	33.3	29.4	57.1	18.9	33.2
ADF	27.1	23.6	20.8	38.8	28.1
HEM	23.5	20.7	12.9	14.7	29.4
CEL	23.3	14.9	15.5	24.5	23.5
LIG	3.3	8.2	4.7	12.5	4.3
TDN	71.8	72.1	74.9	63.9	70.6

¹ DM = Dry matter; CP = crude protein; NDIN = neutral detergent insoluble nitrogen ($g\ kg^{-1}$ of total nitrogen); ADIN = acid detergent insoluble nitrogen ($g\ kg^{-1}$ of total nitrogen); EE = ethereal extract; TC = total carbohydrates; NDF = neutral detergent fiber; NDFap = neutral detergent fiber corrected for ashes and protein; NFC = non-fibrous carbohydrate; NFCap = non-fibrous carbohydrate corrected for ashes and protein; ADF = acid detergent fiber; HEM = hemicellulose; CEL = cellulose; LIG = lignin; TDN = total digestible nutrients.

Table 3. Nutrients intake of mixed-breed male lambs fed experimental diets¹.

Item ²	Treatment ³					CV ⁴ (%)
	Sorghum silage	Pineapple	Banana	Mango	Passion fruit	
<i>Kg day⁻¹</i>						
DM	1.0	1.2	1.1	1.1	1.2	19.8
OM	0.8	1.1	1.1	1.0	1.1	19.8
CP	0.2	0.2	0.2	0.2	0.2	19.3
EE	0.03	0.03	0.04	0.04	0.04	33.8
NDF	0.3	0.2	0.2	0.3	0.4	35.4
NDFap	0.2	0.2	0.2	0.2	0.3	22.1
NFC	0.4	0.6	0.5	0.5	0.4	25.0
NFCap	0.4	0.5	0.5	0.5	0.4	17.8
TDN	0.7	1.0	0.9	0.9	0.8	19.0
<i>% BW</i>						
DM	3.1	3.7	3.4	3.4	3.7	15.8
OM	2.5	3.4	3.2	3.1	3.2	14.7
NDF	0.9	0.7	0.7	0.9	1.1	32.0
NDFap	0.7 ^{ab}	0.7 ^{ab}	0.6 ^b	0.6 ^b	0.9 ^a	18.6
TDN	2.2	2.9	2.7	2.6	2.6	17.0

¹Experimental diets consisted of sorghum silage (control diet) and 75% replacement of sorghum silage for dehydrated fruit processing by-products.

²DM = Dry matter; OM = organic matter; CP = crude protein; EE = ethereal extract; NDF = neutral detergent fiber; NDFap = neutral detergent fiber corrected for ashes and protein; NFC = non-fibrous carbohydrate; NFCap = non-fibrous carbohydrate corrected for ashes and protein; TDN = total digestible nutrients.

³Means followed by different superscript letters in line are statistically different by Tukey test at 5% level of probability.

⁴CV = coefficient of variation.

Table 4. Apparent digestibility of DM, OM, CP, NDFap, NFCap, and TDN of mixed-breed male lambs fed experimental diets¹.

Item ² ($g\ kg^{-1}$)	Treatment ³					CV ⁴ (%)
	Sorghum silage	Pineapple	Banana	Mango	Passion fruit	
DM	71.0	69.2	66.4	69.5	65.3	5.4
OM	72.9	70.8	68.5	70.1	65.7	5.3
CP	68.3	65.9	60.0	66.1	67.0	8.2
EE	73.8 ^a	75.4 ^a	47.9 ^b	65.8 ^{ab}	76.0 ^a	18.0
NDFap	53.5 ^a	39.5 ^{ab}	38.3 ^{ab}	36.3 ^b	28.9 ^b	21.4
NFCap	85.3	85.6	83.3	87.1	86.3	2.8
TDN	69.9	68.2	66.8	66.7	63.4	19.0

¹Experimental diets consisted of sorghum silage (control diet) and 75% replacement of sorghum silage for dehydrated fruit processing by-products.

²DM = dry matter; OM = organic matter; CP = crude protein; EE = ethereal extract; NDFap = neutral detergent fiber corrected for ashes and protein; NFCap = non-fibrous carbohydrate corrected for ashes and protein; TDN = total digestible nutrients.

³Means followed by different superscript letters in line are statistically different by Tukey test at 5% level of probability.

⁴CV = coefficient of variation.

Table 5. Urinary excretion of purine derivatives and microbial efficiency of mixed-breed male lambs fed experimental diets¹.

Item	Treatment ²					CV ³ (%)
	Sorghum silage	Pineapple	Banana	Mango	Passion fruit	
Allantoin (mmol day ⁻¹)	10.9	14.6	17.7	14.9	13.3	34.2
Xanthine-hypoxanthine (mmol day ⁻¹)	1.3	1.9	2.4	2.7	3.4	65.8
Uric acid (mmol day ⁻¹)	1.0	0.9	1.2	0.7	1.0	38.1
Allantoin (<i>g kg⁻¹</i>)	81.9	83.3	82.7	80.9	74.5	10.8
Xanthine-hypoxanthine (<i>g kg⁻¹</i>)	10.3	11.3	11.3	14.9	19.6	64.8
Uric acid (<i>g kg⁻¹</i>)	7.8	5.4	5.9	4.2	5.9	36.5
Total PD (mmol day ⁻¹)	13.2	17.4	21.3	18.4	17.7	30.1
Absorbed PD (mmol day ⁻¹)	15.0	19.9	24.7	21.4	20.5	31.11
Microbial nitrogen (<i>g day⁻¹</i>)	9.2	12.3	15.2	13.2	12.6	31.1
Microbial efficiency (<i>g microbial N/kg TDN</i>)	83.6	80.6	110.8	101.2	93.3	37.6
Serum urea N (<i>mg dL⁻¹</i>)	25.9 ^a	17.3 ^b	18.7 ^{ab}	16.6 ^b	18.2 ^b	19.5

¹Experimental diets consisted of sorghum silage (control diet) and 75% replacement of sorghum silage for dehydrated fruit processing by-products.

²Means followed by different superscript letters in line are statistically different by Tukey test at 5% level of probability.

³CV = coefficient of variation.

Table 6. Nitrogen balance of mixed-breed male lambs fed experimental diets¹.

Item	Treatment ³					CV ⁴ (%)
	Sorghum silage	Pineapple	Banana	Mango	Passion fruit	
Nitrogen intake						
g/day	27.4	34.7	32.6	34.3	32.6	19.3
g/BW ^{0.75}	2.3	2.8	2.8	2.9	2.7	13.6
Fecal nitrogen ²						
g/day	8.6	12.8	15.1	12.5	11.9	31.8
g/BW ^{0.75}	0.7	1.0	1.3	1.0	1.0	22.4
Urine nitrogen ²						
g/day	4.2	3.6	3.2	3.3	2.6	28.2
g/BW ^{0.75}	0.4	0.3	0.3	0.3	0.2	30.6
Nitrogen balance						
g/day	14.7	18.3	14.2	18.6	18.2	23.3
g/BW ^{0.75}	1.3	1.5	1.2	1.6	1.5	22.5

¹Experimental diets consisted of sorghum silage (control diet) and 75% replacement of sorghum silage for dehydrated fruit processing by-products.

²Nitrogen excreted from feces and urine respectively.

³Means followed by different superscript letters in line are statistically different by Tukey test at 5% level of probability.

⁴CV = coefficient of variation.